Appendix 3

1. Paradigms

Experimental paradigms used in RM's articles are listed in **Table 1** with: (i) the description of the paradigm; (ii) an abbreviation for the paradigm used in **Table 4**; (iii) how the paradigm is referred to by RM in the title, method, results, or figures in different articles; and (iv) the number of articles using the paradigm. In RM's articles, to the best of my understanding, predator odor is both paradigm and method, so it is listed in both **Table 1** with paradigms and **Table 2** with methods

.

Table 1. Paradigms			
Experimental paradigm or group	**Abbreviation**	**How it is referred to in the data (figures, methods, results, title)**	**Number of articles used**
Environmental enrichment 14 – 16 days	E	*Environmental enrichment*	6 out of 8 articles
		Short-term environmental enrichment	
		Early-life short-term environmental enrichment	
		Complex housing	
		Complex environment	
		Enriched environment	
		Enriched	
		Complex	
Chronic immobilization stress 2 hours for 10 days started same day as enrichment	S	*Stress*	3 out of 8 articles
Control	C	*Animal facility rearing (ARF)*	7 out of 8 articles
		Standard	
		Not enriched	
		Non-enriched	
		Simple	
		Control	
Maternal Separation PN2-14 once/day for 3 hours	M	*Stress*	3 out of 8 articles
		Pre-weaning stress	
		Anxiety	
		Stress-induced anxiety	
		Maternal separation	
Predator odor	P	*Predator odor exposure*	2 out of 8 articles
		Predator stress	
		Stress	
		Stressor	
Deep Brain Stimulation	D		1 out of 8 articles
Toxoplasma gondii	T	*Infected*	1 out of 8 articles

2. Methods

RM's articles obfuscate apparatus, methods, results, and data interpretation. In **Table 4** methods are referred to and grouped as they are generally understood. As such, the methods and an abbreviation for each are listed in **Table 2** next to terms and variations used in RM's articles for the same. The text in italics in **Table 2** is copied from RM's articles. This text refers to a method or test, and a definition of the readout as given by RM. To the best of my ability, the readout as finally reported by RM in an article is in **Table 4**. The list of terms and definitions for a test used by RM in **Table 2** is not exhaustive and does not include variations of interpretations of those tests in RM's articles, for example in the Discussion of those articles.

Table 2. Experiments

Method	Abbreviation	RM nomenclature or operational definition	Comment
Behavior			
Home cage emergence	HCE	*Latency to leave the cage*	
		Latency to emerge from the home cage	
		Latency to emerge from the home cage into a novel environment	
		Stress-induced anxiolysis	
Object Recognition Task	ORT	*Object recognition task*	Used in one study.
		Memory performance	
		Novel object recognition	
		Exploration of novel and familiar objects was quantified over a period of 180 s. Object recognition was quantified as exploration of novel objects relative to the sum of exploration for novel and familiar objects	
Elevated Plus Maze	EPM	*Relative open arm exploration*	This is consistently referred to as EPM by RM across articles, but I cannot comment on why the common and simple readouts were not used for this test, nor if RM's operational definitions are the same. The readouts are different from one article to another, listed to the best of my ability in **Table 4**.
		Exploration in open and enclosed arms was quantified for five minutes each	
		Open arm exploration (entries and occupancy time) relative to the sum of open and enclosed arm explorations was used as an index for anxiety. Mean of percentage open arms entries and percentage open arms time was subtracted from 100 to derive an index for anxiety	
		Manually analysed to quantify the percentage open-arm time and open-arm entries, relative to the sum of open and enclosed arm exploration / with decreased open-arm entries and time indicative of heightened anxiety	
		Open arm exploration (entries and occupancy time) relative to sum of open and enclosed arm exploration was used as an index for anxiety	
		Open arm exploration in an elevated plus maze (EPM) was studied as an index of anxiety (lower exploration = greater anxiety). It was defined as mean of % open arm entries and % open arm time (relative to sum of open and enclosed arm entries and time)	
		general locomotion tested on the EPM	
	Head dips	*Risk assessment*	Separate sections in articles and broad interpretations are based on this. It is still EPM.
		Exploratory behavior	
		Goal-directed behavior	
		Absence indicates passive behavior	

Method	Abbreviation	*RM nomenclature or operational definition*	Comment
Behavior			
Open Field Test	OFT-R/C	*Open field test*	This is reported square in some articles, circular in others denoted in **Table 4** by OFT-R or OFT-C respectively. OFT-C is reported big or small as detailed in *Appendix2.pdf.*
		Open field	
		Arena	
		Novel rectangular arena	
		A square-shaped open field was constructed / Time spent in the central part of the open field (33 cm x 33 cm) was measured as a proxy for lower anxiety-like behavior	
		Time spent in the centre of the field was quantified as the reciprocal proxy of the anxiety (centre defined as a concentric circle to the arena with 0.33 m radius). Total distance travelled during the trial was also quantified as a measure of locomotion	
		Time spent in the center of the arena was quantified from recorded videos. The center was defined as a concentric circular region of interest with a radius of 0.33 m. In a subset of animals, total time spent by animals being mobile in the arena was also quantified as an index of locomotion	
Predator odor	PO-R/C	*Predator odor exposure*	This is used as a stress paradigm as indicated in **Table 1** in some articles, as a method in other articles. Predator odor exposure in a rectangular arena is denoted by PO-R, and circular by PO-C.
		Predator stress	
		Stressor	
		Open field	
		Open field arena	
		Maze	
		Rectangular arena	
		Circular arena	
		Maze	
		Innate aversion	
		Occupancy of the bisect containing odor relative to the total area of the arena (chance = 47.2%, based on the area of the bisect containing predator odor vis-à-vis total area of the arena)	
		Aversion to cat odor was quantified as percentage time in bisect containing cat odor relative to total trial duration	
		Center time in open field	
		Percentage time in bisect containing cat odor relative to total trial duration	
Golgi staining			
Dendritic Length	DL	*Total dendritic length*	
Dendritic Branching Points	DB	*Total number of branch points*	
		Number of branch points	
		Dendritic arborization	
		Dendritic complexity	
		Pyramidal or modified pyramidal spiny projection neurons	
		Sholl analysis	

Method	Abbreviation	*RM nomenclature or operational definition*		Comment
Golgi staining				
contd. Dendritic Branching Points	DB	*Primary dendrites*		
		Mean of these dendritic segments on different neurons was used to estimate spine density for each animal		
		Number of primary dendrites was estimated by counting dendritic intersection with a circular region of interest (radius = 30 µm) centered on the cell soma. A Sholl analysis was further conducted for PrL neurons by overlaying multiple circular regions of interest centered on the cell soma and successively increasing the radius in step size of 10 µm. Number of intersections made by the dendritic arbors was then quantified as a function of radial distance from the soma		
		Dendritic length and number of branch points were quantified as function of radial distance from the cell soma		
		Dendritic architecture of PrL neurons was further analyzed as a function of radial distance from soma using Sholl analysis		
Dendritic spines	DS	*Spine density*		Manually counted
		Spine count		
		Spine density per 60 µm		
		Scholl's both total branch points and segmental branch points at radial distance		
		Primary and secondary dendrites		
		Primary dendrites	*Dendrites directly originating from the cell soma*	
			spine density (1/100 µm) counted along 80 µm stretch of dendrite	
			spine density (1/100 µm) counted along 60 µm stretch of dendrite	
		Secondary dendrites	*the first branch emerging from the primary dendrite*	
			those originating from primary dendrites	
			spine density (1/100 µm) counted along 80 µm stretch of dendrite	
			spine density (1/100 µm) counted along 60 µm stretch of dendrite	
Dendritic Intersections		*Dendritic intersections*		Though reported, I cannot find it.
		Segmental branch points		
		Sholl analysis		
Brain region				
Basolateral amygdala	BLA			
Prelimbic cortex	PL	*Prelimbic medial prefrontal cortex*		
		Prelimbic cortex		
		Prelimbic region of medial prefrontal cortex		
Infralimbic cortex	IL	*Infralimbic region of medial prefrontal cortex*		
Medial prefrontal cortex	mPFC			
Hippocampus	HP			

Method	Abbreviation	*RM nomenclature or operational definition*	Comment
Other			
Corticosterone	CORT		These are mentioned here for abbreviations in **Table 4**.
Enzyme-linked immunosorbent assay	ELISA		
Glucocorticoid receptor	GR		
Mineralocorticoid receptor	MR		
Brain-derived neurotrophic factor	BDNF		
Polymerase chain reaction	PCR		
Immunohistochemistry	IHC		
Western blot	WB		

3. Article data

Table 4 summarizes article paradigm, method and readout, statistics, significance and effect size reporting, and unknown data in RM's publications. All text in italics in **Table 4** is quoted from the articles. This includes statistics used, the descriptive text is abridged from the articles. **Table 3** is a legend for **Table 4**.

- **Animals:** all results reported in all experiments were from adult male Wistar rats 6 to 8 weeks, except in Abdulai-Saiku et al. 2017, the sex is denoted in **Table 4** along with paradigm by ♂ or ♀.
- **Euthanasia:** when mentioned, animals were killed by decapitation and/or cardiac perfusion. This is related to present work and RM's AUP A:19027. See **Table 6**.
- **Articles:** those are the same as discussed in *Appendix2.pdf*, primary articles (not reviews) produced by RM at NTU.

Table 3. Legend

Article Paradigm	**Method**	**Readout reported**	**n (significance)**	**Statistics reported, figure data**	**Unknown**
			Paradigm 1/paradigm 2/etc. (C: control) **Number of animals used in a paradigm (or neurons)** **Significance reporting**		
Article reference **Paradigm**	Method 1	*Readout (article text)*	X/X/X/X	*Abridged text from article on statistics and figure data reporting if present.*	Non-exhaustive list of: - Unreported methods or results. - Information mentioned said to be reported in part of the article but not found. - Methods information mentioned in another article by RM but not in this one. - Contradictory information in the article.
	Method 2	*Readout 1*	X/X/X/X		
		Readout 2	X/X/X/X		
		Readout 3	X/X/X/X		
		Legend			
		Numbers in red indicates statistical significance reported in Figure	X/X/X/X		
		Numbers in blue indicates effect size reported in results (not figure)	X/X/X/X		
		Numbers highlighted in yellow indicates contradictory reports of significance or effect size in results and figure	X/X/X/X		
		Text highlighted in green indicates data reported in results but not methods, or methods but not results	X/X/X/X		

Table 4. Data					
Article Paradigm	**Method**	**Readout reported**	**n (significance)**	**Statistics reported, figure data**	**Unknown**
			CC/MC/EE/ME		
Koe et al. 2016 **E, M**	HCE	*Emergence latency*	12/12/11/14	*Normality using Shapiro-Wilk; non-parametric for intergroup comparisions (Kruskal-Wallis one-way analysis of variance); Mann-Whitney U 2-tailed if significant intergroup; C vs. MS ± EE; resultant Type 1 errors adjusted with Bonferroni; effect with non-parametric Cliff's delta.* *Figures depict median and interquartile range except for DL and DS average value/animal/n unique neurons*	- Timing of OFT (age of rats at time of OFT) - Objective lens interface - Coordinates for histology - Software for analyzing distance travelled - Calibrated scale for drawing tube scans - Neuron sampling method - Neuron data analysis script
	EPM	*Open arm entries (%)*	14/10/15/10		
		Time spent in open arms (%)	14/10/15/10		
		Number of head dips	14/10/15/10		
	OFT-C	*Time spent in open field center*	14/10/15/10		
		Distance travelled / locomotion	?/?/?/?		
	DL-BLA	*Total dendritic length 10-11 or 8-11 neurons/animal*	6/9/9/9		
	DB-BLA	*Total number of branch points 10-11 or 8-11 neurons/animal*	6/9/9/9		
	DS-BLA	*Spine density 6-10 neurons/animal/60 µm neuron, primary*	6/9/9/9		
		Spine density 6-10 neurons/animal/60 µm neuron, secondary	6/9/9/9		
	DL-PL	*Total dendritic length 8 neurons/animal*	6/9/9/9 4/10/7/9		
	DB-PL	*Total number of branch points 8 neurons/animal*	6/9/9/9 4/10/7/9		
	CORT-ELISA	*Serum corticosterone*	9/9/9/9		
			CC/SE/EE/SE		
Ashokan et al. 2016 **E, S**	EPM	*% open arm exploration (%)*	17/6/8/8	*2-way ANOVA (inter-subject source of variance = stress and EE); planned comparision for stress ± EE (± EE); two orthogonal comparisions Sidak's multiple comparision.* *Figures represent mean and SEM and mean inter-group differences and SEM.*	- Number of animals for results - Kit used for corticosterone analysis - Mean inter-group differences and SEM - Neuron sampling method - Objective lens - Method of Scholl analysis - Neuron data analysis script
		Enclosed entries	17/6/8/8		
		Number of head dips	17/6/8/8		
	DL-BLA	*Dendritic length*	8-7/7/7/7		
	DB-BLA	*Number of branch points 8-10 neurons per animal*	8-7/7/7/7		
		Number of branch points vs. distance from soma	8-7/7/7/7		
	DS-BLA	*Spine density per 60 µm neuron Primary and secondary simultaneously*	8-7/7/7/7		
	CORT-ELISA	*Corticosterone measurement*	8/7/6/6		
	BDNF-PCR	*BDNF mRNA abundance*	4/4/4/4		
	Adrenal wt.	*Adrenal weight*	?/?/?/?		

Article Paradigm	Method	Readout reported	n (significance)	Statistics reported, figure data	Unknown
			P		
Hegde et al. 2017 **P**	EPM	*% Open arm time*	9	*Jackknife resampling model; Spearman's rank correlation coefficients for pairs of endpoints; principal component analysis; dimension reduction; factor loading exceeded/less 0.5 for component/alternative; 4/3 endpoints load on the 1st/2nd principal component.* *Figures depict raw data median %/unit/principal component*	- All data from control group (n=6) - Data from predator exposure in rectangular arena - Source of bobcat urine - Objective lens - Neuron data analysis script
		% Open arm entries	9		
		Number of enclosed arm entries	9		
	PO-R	*Occupancy of bisect containing odor relative to the total area 47% chance*	?		
	OFT-R	*Center time in open field*	10		
	DB-BLA	*Median number of branch points*	10		
	DL-BLA	*Median dendritic length*	10		
	DS-BLA	*Primary and secondary dendrites* *Median spines on primary dendrites*	8		
	CORT-ELISA	*Corticosterone: post-open field, post-EPM, post-sacrifice*	10		
			♂C/M♂		
Abdulai-Saiku et al. 2017 **P, M, T**	EPM	*Entries in open arms (%)*	8/8	*Probability type 1 error unpaired two-tailed Student's t-test; standardized effect size Cohen's d; values magnitude 0.8 robust; negative d correspond comparisons mean of respective controls; mean intergroup 95% confidence intervals; univariate analysis of variance % open arm entries employing number of enclosed entries as a covariate.* *Figures presented as mean SEM, along with individual values for each animal* *for each endpoint*	- Source of cat urine - Mean and SEM - Method and data for serological exam
		Time in open arms (%)	8/8		
		Head dips	?/?		
	HCE	*Escape latency (s)*	8/8		
			♀C/♀P		
	PO-R	*Time spent in cat bisect (%)*	8/7		
	PO-C	*Time spent in cat bisect (%)*	10/9		
	?	*Serological exam*	?		
			CC/SC/EE/SE		
Ashokan et al. 2018a **E, S** contd.	Forced swimming test x2 days	*Time spent immobile (s) (trial 2)*	8/8/8/6 or 8	*2-way ANOVA to estimate main effects; orthogonal planned comparisons between stressed and non-stressed in either enriched or non-enriched using independent sample t-test.; planned comparisons guided by a priori interest; non-enriched and enriched groups in absence of stress not compared using planned comparisons; eta squared.* *Figures represent mean and SEM.*	- Primary antibody catalogue - Secondary antibody for DCX - Number of sections for BrdU and DCX - Combining Golgi and IHC method - Device for measuring immobility data - Neuron data analysis script - First trial swimming test - Number of sections for counting BrdU - Lens objective magnification
		Latency to immobility	?		
		Climbing	?		
		Number of immobility events/episodes (Trial 2)	8/8/8/6 or 8		
	?	*Baseline immobility - device*	?		
	OFT-R	*Baseline immobility,* *Total number of sorties*	?		
		Baseline immobility, *Total time spent immobile (s)*	8/8/8/8		
		Baseline immobility, *Total distance travelled*	?		

Article Paradigm	Method	Readout reported	n (significance)	Statistics reported, figure data	Unknown
contd.	DS-HP	*Total number of spines on primary dendrite across 60 μm in CA3, 2 neurons /animal/group (spine density /60 μm)*	50/49/38/43		- Calibrated scale for drawing tube scans
Ashokan et al. 2018a	BrdU HP-ventral	*Enumerated in 10 sections, 4-6 animals*	?		
	BrdU HP-dorsal	*Enumerated in 10 sections, 4-6 animals, dorsal dentate gyrus/CA3*	4/5/5/6/		
E, S	Doublecortin	*Enumerated in 8 sections for each animal*	?		
	GR-PCR	*PCR real-time fluorescence Raw values abundance mean*	4/4/4/4		
	MR-PCR	*PCR real-time fluorescence Raw values abundance mean*	3/4/4/4		
			C/E		
Ashokan et al. 2018b	DL-PL	*Total dendritic length (μm)*	8/4	*Mean for each animal, across multiple neurons analyzed; unpaired two-tailed Student's t-test; standardized effect size using Cohen's d values above magnitude of one robust; negative d values correspond to the comparisons where mean of complex housing greater than simple housing; mean inter-group difference 95% confidence intervals; repeated measure analysis of variance for Sholl analysis for number of intersections as a function of radial distance from the soma. Data presented as mean SEM along with individual values.*	- Enrichment protocol - Objective lens magnification - Neuron data analysis script - Calibrated scale for drawing tube scans
		Total number of intersections Sum of intersections (#)	8/4		
	DB-PL	*Primary branches (#)*	8/4		
		Maxima for the number of the intersection, maximum intersections (#)	8/7		
		Radius for the maxima of the intersection, radius max intersection (μm)	8/7		
		Intersections (#)	8/7		
E	DS-PL	*1° spine density (1/100 μm) counted along 80 μm stretch of dendrite*	8/4		
		2° spine density (1/100 μm) counted along 80 μm stretch of dendrite	8/4		
	DL-IL	*Total dendritic length*	6/6		
	DB-IL	*Total number of intersections Sum of intersections*	6/6		
	DB-IL	*Primary branches (#)*	6/6		
	DS-IL	*1° spine density (1/100 μm) counted along 60 μm stretch of dendrite*	6/6		
		2° spine density (1/100 μm) counted along 60 μm stretch of dendrite	6/6		

Article Paradigm	Method	Readout reported	n (significance)	Statistics reported, figure data	Unknown
			CC/DC/EE/DE		
Bhaskar et al. 2018 **E, D**	HCE	*Ranks, home cage emergence*	10/9/10/8	*Normality Shapiro-Wilk test significant departure from normality; nonparametric statistics intergroup comparisons Kruskal–Wallis; each endpoint rank-transformed across four groups; two planned comparisons set before data collection started; orthogonal comparisons, no group more than one comparison; independent sample Student's t for parametric planned comparisons of rank transformed data; planned comparisons buttressed by effect size data using Cohen's d*	- Power source for deep brain stimulation - Some parameters of deep brain stimulation - Values for individual animals - Mean and SEM for figures
	EPM	*Ranks, EPM anxiety index*	10/9/10/8		
		Ranks, head dips	10/9/10/8		
	ORT	*Ranks, novel object recognition*	10/9/10/8		
.			**CC/MC/EE/ME**		
Hegde et al. 2020 **E, M**	OFT-C	*Time in center (s)*	12/8/8/6	*Outliers removed using ROUT method with a max false discovery rate 1%27; main effects estimated using a two-way analysis of variance; omnibus post-hoc comparisons not conducted to avoid multiple pair-wise contrasts; two orthogonal planned comparisons constructed using independent sample t-test between non-stressed controls and stressed animals under animal-facility rearing and between non-stressed controls and stressed animals reared in enriched housing*	- Softwares for behavioral analysis and image analysis - Number of animals for neuron analysis and IHC - Primary and secondary antibodies - Objective lens -Calibrated scale for drawing tube scans - Assessment for GR promoter methylation.
		Time mobile (s)	6/5/9/6		
	CORT-ELISA	*Baseline corticosterone*	6/5/9/6		
	Adrenal wt.	*Wet adrenal weights mg/kg bd. wt.*	6/5/9/6		
	DS-BLA	*Spine density* *Averaged primary secondary 60 µm*	60/60/60/60		
	GR-IHC	*Intra-nuclear GR*	24/22/24/24		
	GR-WB	*GR relative intensity AU*	6/6/6/6		
	GR-?	*GR promoter methylation*	5/5/5/5		
	BDNF-IHC	*BDNF rise constant AU*	24/24/24/26		
		BDNF Y max AU	22/21/24/24		
	BDNF-WB	*BDNF relative intensity AU*	6/6/6/6		
Wu and Mitra 2020	This article violates known ethical, physiological, and physical laws and so was excluded from this analysis.				

4. Duplicate publications

Articles with datasets possible to extract from one study are shaded the same color.

Table 5. Duplicate publications

Acknowledgement	RM's funding	RM's paper	AV's paper	Shared authors	AV's funding	Comments
Ling, NY	NTU M4081146	***Koe et al., 2016***				
Vyas, A						
Bhaskar, Y	NTU	***Ashokan et al., 2016***				
Vyas, A	MoE RG 46/12					
Ashokan, A	NTU	***Hegde et al., 2017***				
Vyas, A	MoE RG46/12					
	MoE RG136/15	***Abdulai-Saiku et al., 2017***	***Singh et al., 2020***	Abdulai-Saiku, A	MoE RG136/15	The methods are incomplete, but there is a typo: testosterone cannot be dissolved to 25 mM in absolute ethanol (3% reported). Maybe it's 25 μM.
	MoE RG46/12		***Abdulai-Saiku and Vyas, 2017***	Abdulai-Saiku, A		There might be a readout novel to RM for PO-R (Entries into bisect) in AV's paper.
	MoE RG 46/12 to RM	***Bhaskar et al., 2018***	***Liu et al., 2015***	Lim, LW	NTU, Lee Wei Lim, Lee Kuan Yew Research Fellowship M4080846.080	
Koe, A	MoE RG 46/12	***Ashokan et al., 2018a***				
Vyas, A	NTU					
	MoE RG 46/12 to RM	***Ashokan et al., 2018b***				
Vyas, A	MoE RG 144/17 to RM	***Hegde et al., 2020***				
			Tan et al., 2015	Lim, LW	MoE: DT, LJTS, LWL, AV	Are these different studies?
					Duke-NUS block funding: TCWD, XZ	
			Tan and Vyas, 2016a		MoE RG52/14 to TD and AV	
			Tan and Vyas, 2016b		MoE RG52/14	
			Tong et al., 2019	Abdulai-Saiku, A	MoE RG136/15	These two articles look quite similar.
			Tong et al., 2020	Abdulai-Saiku, A	Human Frontier Science Program RGP0062/2018	

5. AUP: A19027

Table 6. AUP: A19027		
Section	**RM**	**Comment**
6a. Avoiding duplication	"…We do not initiate any project unless it is sufficiently novel, first because it will not be published since it cannot add any new information to the field, second, **do not need/plan to use animals just to check something already published and replicated before**. Thus when we initiate a project the maximum value is in its novelty **where most of the work is likely first time, with some replicability options**. Since our work is about animal models of psychopathology, measure of behavioural output is crucial. However **the final read-out** of projects also critically involve other important measures of physiology, protein and gene regulation. So there is minimal overlap between the main questions of different projects, hence minimising duplication of earlier work…" (emphasis added)	Please see **Table 4**.
6e. Reduction	"…I have a good approximation of the number of animals needed for optimal behavioural output… we calculated approximate number of animal required through statistical power analysis…"	"…Please bring down the increase in animal numbers to less than 20% of the current approved animal numbers, which is less than 180 mice to qualify as a minor amendment…" [For Revision, ICCO, 28-Aug-2020]
Description of non-surgical procedures	"…The lack of use of anesthesia for decapitation is important in experiments (described in part 11) where mRNA levels are measured/ visualized. The animal when presented an odor, will process the sensory information in specific areas of the brain that will elicit a change at the gene expression level- either turning on some genes or turning off other genes. Thus, the use of anesthesia can be a confounding factor as this will also elicit its own changes at the gene expression level. Hence, it would be difficult to separate the effect of the odor and anesthesia. To reduce pain and discomfort during this procedure, Harvard restraint bags will be used to hold down the animal and a quick and smooth decapitation (taking less than a minute that also serves to reduce discomfort) with the guillotine will be done…"	I had been hoping RM neglected to mention anesthetic overdose for killing animals. In light of evidence presented here, notably **Tables 1, 4** and **5**, I think this might not be the case.
12b. Genetically Modified Organisms (GMOs) (i) Is Recombinant DNA or transgenic animal used? GMAC approval must be obtained and attached to this section tab.	"…No…"	I do not think this is true.

6. References

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4. Ashokan, A., Hegde, A., Balasingham, A., and Mitra, R. (2018a). Housing environment influences stress-related hippocampal substrates and depression-like behavior. *Brain research* **1683**, 78-85. https://www.sciencedirect.com/science/article/abs/pii/S0006899318300350 10.1016/j.brainres.2018.01.021

5. Ashokan, A., Lim, J.W.H., Hang, N., and Mitra, R. (2018b). Complex housing causes a robust increase in dendritic complexity and spine density of medial prefrontal cortical neurons. *Scientific reports* **8**, 1-9. https://www.nature.com/articles/s41598-018-25399-4 10.1038/s41598-018-25399-4

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